

Journal Pre-proof

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PII: S2352-1864(19)30750-3
DOI: <https://doi.org/10.1016/j.eti.2020.100675>
Reference: ETI 100675

To appear in: *Environmental Technology & Innovation*

Received date: 31 October 2019
Revised date: 31 January 2020
Accepted date: 9 February 2020

Please cite this article as: L.K. Pandey, In situ assessment of metal toxicity in riverine periphytic algae as a tool for biomonitoring of fluvial ecosystem. *Environmental Technology & Innovation* (2020), doi: <https://doi.org/10.1016/j.eti.2020.100675>.

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1 **In situ assessment of metal toxicity in riverine periphytic algae as a tool for biomonitoring**
2 **of fluvial ecosystems**

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11 **Keywords-** Chemical diffusing substrates; Diatoms; Lipid bodies; Deformities; Heavy metal
12 toxicity; Biomonitoring; Ecotoxicology

13 **Abstract**

14 The combined effects of seasonality and in situ heavy metal (Cu, Zn, Pb) enrichment on
15 periphyton in a large river was experimentally studied using metal-diffusing substrates. Highest
16 release rate and intracellular accumulation of metal ions (Cu, Zn and Pb) was examined in rainy
17 and summer seasons, respectively. Release rate and intracellular accumulation showed the
18 following order: Zn>Cu>Pb in the three seasons. Periphytic algae growing on these substrates
19 showed intracellular accumulation of test metals (Cu, Zn and Pb) which inhibited growth in a
20 concentration dependent manner, as evidenced by reduced cell number, species richness and
21 Shannon index of the community. Concentration dependent decrease in Chlorophyll *a* and rise in
22 Caro./Chl *a* ratio was found under Cu and Zn stress. Periphytic community in summer and

23 winter was found similar than rainy season and was found be dominated by diatoms followed by
24 green and cyanobacteria. In comparison to green algae and cyanobacteria, diatom species,
25 showed relatively higher % relative abundance under heavy metal stress than their respective
26 control. Increased lipid body production and cell wall deformities in diatoms were found under
27 Cu and Zn stress but not under Pb stress, possibly because Pb diffusion from the substrates was
28 low. After 28 days of diffusion, metal release rates were near the control levels and periphyton
29 parameters had recovered, demonstrating rapid response to changing conditions by the
30 periphyton. The rapid recovery of the periphyton (within 14 days) indicates that periphyton
31 biomonitoring may be useful for not only monitoring of stable conditions but also for changing
32 conditions, such as monitoring recovery. The present study shows the utility of metal diffusing
33 substrate and periphytic diatom community as an effective tool for biomonitoring (including
34 recovery) of heavy metal pollution in the fluvial ecosystem.

35 **1. Introduction**

36 Metal toxicity to periphyton has been often studied in polluted habitats, like acid mine drainage,
37 metal smelter waste waters, and metal contaminated waterbodies (Morin et al., 2008a, b; Pandey
38 et al., 2016). In many of these studies, carried out mostly on fluvial systems, periphyton of metal
39 impacted stations have been compared with those from the reference (unpolluted) station to get
40 an insight into how metal pollutants affect this particular community as also how the community
41 recovers after a certain distance downstream. Artificial attachment sites have been often used so
42 as to provide uniform substrates for overcoming variability in natural substrates found in a
43 waterbody. Glass slides, with rough surface to encourage periphytic colonization, has been the
44 common choice of periphyton ecologists (Weitzel, 1979). Periphytic species richness, diversity,
45 similarity and even multivariate tools have often been related with metal concentration in water

46 (Pandey et al., 2016, 2017). Nevertheless, such studies do not give us an idea about how
47 periphyton of an unpolluted or pristine aquatic system would respond in case of metal
48 enrichment. Another disadvantage is that one cannot be certain if the observed effect is due to
49 metal or some other environmental perturbation. For instance, flow conditions in running water
50 environments are highly variable and hence can have tremendous impact on algal periphyton
51 (Ghosh and Gaur, 1994): if the control and the metal impacted sites do not have identical flow
52 regimes, we cannot compare periphytic algae of two such sites to derive plausible conclusions
53 with regard to heavy metal toxicity.

54 Efforts have been made to study metal toxicity to periphyton in laboratory streams or outdoor
55 channels (Serra et al., 2009; Pandey et al., 2015). One of the major advantages of such an
56 approach is ease in experimentation. Another advantage is that the researcher is sure about the
57 observed effects being elicited due to metal because appropriate controls are always considered.
58 However, environmental conditions in the laboratory or outdoor channels are vastly different
59 from those occurring in natural bodies of water. This is one of the serious limitations of the
60 artificial channels.

61 Now we are confronted with the question as to how can we study the effect of a toxicant or
62 nutrient on periphyton in situ in a river or lake. Enriching the entire body of water with the test
63 chemical is not an environment-friendly proposition as it may disturb the ecology of the system.
64 Incidentally, this has been done to prove the critical role of phosphorus in triggering
65 cyanobacterial blooms and eutrophication (Schindler, 1974). A novel way of in situ studying
66 effect of nutrients (such as, nitrogen and phosphorus) on periphyton was developed in the form
67 of nutrient diffusing porous clay substrates (Ghosh and Gaur, 1994; Scott et al., 2009). In this
68 kind of set up, there is no need to add nutrients to waterbody; rather nutrients diffusing out of

69 porous clay substrate can be directly taken up by periphyton growing onto them and this may
70 facilitate studying the effect of in situ nutrient enrichment on periphyton.

71 Prompted by earlier studies on nutrient diffusing substrates, as mentioned above and the
72 work of Arnegard et al. (1998), a novel metal diffusing clay substrate was prepared (Pandey et
73 al., 2014; Pandey and Bergey, 2018) and used to in situ study the effect of zinc, copper and lead
74 on periphyton community of a river. The study was carried out in summer, rainy and winter
75 seasons. For this purpose, artificial substrates were filled with metal solutions to assess their
76 effects on periphyton. The artificial substrate could substantially release metal ions from its
77 porous surface for studying the effect of metal toxicity to periphytic community.

78 **2. Materials and methods**

79 *Study area*

80 The study was carried out in the river Ganga at Varanasi (25° 18' N and 83° 1' E; 82 m
81 above m.s.l.). The study area lies in the Indo-Gangetic plains and is characterized by tropical
82 climate greatly influenced by monsoon. During the course of the study, the atmospheric
83 temperature was very harsh during winter (November to February; 6-24 °C during the coldest
84 month January) and summer (March to June; 29-45°C during the hottest month May); rainy
85 season (end of June to October) was hot (33–43°C during July) and humid. The annual total
86 rainfall of Varanasi is ~1100 mm. Several sites were surveyed to find out one which is relatively
87 not much influenced by human activities and where artificial substrates can be placed without
88 any incident of vandals disturbing them. Another criterion for the selection of site was
89 approachability and ease in sampling. Keeping all these points in mind, a small stretch of the
90 river was identified near the campus of the Banaras Hindu University, in Garhwa ghat (Ramna)
91 area, which is very close to the new bridge spanning the river Ganga. The site selected for the

92 study lies before the main city and hence it is relatively free of pollution. At the study area, the
93 width and depth of the river vary seasonally, attaining their maxima during the rainy season. At
94 the place where the substrates were kept, the river was about 2 m deep. The bottom of the river is
95 sandy.

96 Nutrient diffusing clay substrates have been used to in situ assess nitrogen and
97 phosphorus enrichment effect on periphyton of rivers and lakes (Ludwig et al., 2008, Scott et al.,
98 2009). Inspired by these reports, a metal diffusing substrate (MDS) was developed to assess the
99 effect of metal enrichment on river periphyton (Fig. 1). Each MDS was made by fixing a circular
100 porous clay tile (fired in brick kiln; diameter 14 cm and thickness 4 mm) to the wide mouth
101 (diameter 13.5 cm) of a plastic funnel (capacity 670 ml) using an epoxy resin (m-seal; Pidilite
102 Industries, Daman, India). Solutions of copper ($\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$), zinc ($\text{ZnCl}_2 \cdot 5\text{H}_2\text{O}$) and lead
103 (PbCl_2) were prepared in Milli-Q water using their analytical grade salts (Rankem, India). These
104 solutions had 1(low), 2.5 (medium) and 5 (high) g l^{-1} concentrations of each test metal. Metal
105 solution was filled in MDS through the open end of the funnel, which was subsequently closed
106 with a replaceable rubber cork. Metal ions diffused out from numerous tiny pores on the surface
107 of clay tiles when placed in water. The pattern of release of the test metal ions from MDS was
108 determined in triplicate by keeping them in the river. MDS, filled with the selected
109 concentrations of the metal solutions, were placed in the river, as described below, for estimating
110 the rate of release of metal ions. Every week, MDS were sampled and the concentration of metal
111 ion remaining in the solution was measured using an atomic absorption spectrophotometer
112 (PerkinElmer, AAnalyst 800). These data were used to calculate the rate of release of the test
113 metal ions from MDS.

114 Two liters of river water sample were collected at weekly interval in plastic bottles.
115 Collected water was carried to the laboratory within 30 minutes and refrigerated.
116 Physicochemical parameters such as pH (Hanna pHep® tester), conductivity (Milwaukee
117 stainless steel probe) and temperature were directly measured in the river. The flow rate was
118 measured by measuring the time taken by a thermocol float to travel a specified distance. The
119 methods used for various analyses are described below. Nitrate- and nitrite-nitrogen, total
120 phosphorus and dissolved silica was estimated by the methods given in Wetzel and Likens
121 (1979). 50 mL subsample from the collected water was centrifuged at 4500 rpm for estimating
122 metal concentration by using ICP-optical emission spectrometry (ICP-OES, GemCone™
123 nebulizer, dual-view optical system, Perkin-Elmer, Optima 7300 V, USA).

124 The experiment was carried out during three seasons: summer (May-June, 2010), rainy
125 (August-September 2010) and winter (November-December 2010). Three replicates were
126 considered for each treatment and the control. MDS filled with metal solution of concentrations
127 specified above were placed 15 m away from the bank of the river Ganga with their clay tile
128 surfaces lying horizontally about 2 cm below the water surface (Fig. 1). MDS were mounted on
129 bamboo frames that were fixed in the river with the help of bamboos buried vertically deep in the
130 river sediment. The bamboo frames were placed parallel to the direction of river flow because it
131 ensured similar flow conditions for all MDS.

132 **Collection and analysis of river water**

133 *Collection and study of periphyton*

134 Sampling of periphyton was done 7, 14, 21 and 28 days after MDS were placed in the river by
135 scraping ~ 40 cm² area of the colonized porous tile (each time a fresh area). Periphyton samples
136 were collected in glass centrifuge tubes with the help of a blade and stiff brush. Collected

137 samples were taken to the laboratory in ice-packed boxes within 30 min. In the laboratory, these
138 samples were divided into two parts (10 ml each). One part (10 ml) was further divided into two
139 parts (5 ml each), of which one part were quickly subjected to 90% acetone treatment for
140 pigment extraction while the remaining 5 ml was fixed with 4% formaldehyde for identification
141 and enumeration. The second part of the periphytic sample (10 ml) was used for estimating the
142 intracellular accumulation of the test metals.

143 Fresh periphytic samples were also microscopically examined for identification of algae
144 and cyanobacteria often at 450x magnification; higher magnification was used as and when
145 required. Diatom frustules were examined after cleaning. Permanent slides of diatom frustules
146 were prepared as per Pandey et al (2018). The method involved treating periphytic algal samples
147 with 90% acetone and leaving it for one day for removing cytoplasmic content. Subsequently,
148 the samples were dried and treated with concentrated H_2SO_4 till white fumes stopped emanating
149 from the samples. Thereafter, samples were treated with hydrogen peroxide for 30 min, and then
150 washed thoroughly with deionized water. After drying, samples were mounted in Pleurax
151 mounting medium (Refractive index 1.73) onto glass slides for microscopic examination.

152 In the periphyton, individual diatom cells were identified as per Algaebase, AIDI (Pandey
153 et al., 2016) and ANSP (2012) algal image database. In addition, monographs on diatoms by
154 Patrick and Reimer (1966, 1975) and a manual on stream periphyton by Biggs and Kilroy (2000)
155 were also consulted. Coccoid green algae were identified from Phillipose (1959). Cyanobacteria
156 were identified from the monograph of Desikachary (1959).

157 Algal cells were counted to estimate densities with a Spencer's haemocytometer at 450x
158 magnification in 1 ml (n=3) of composite periphyton specimens. Cell density data was used for
159 examining biovolume estimation of the periphytic diatom community as well as for calculating

160 % relative abundance of different algal classes and dominant algal (diatoms, green algae and
161 cyanobacteria) taxa under control and heavy metal stress.

162 The amount of chlorophyll *a* and carotenoids in periphytic samples were determined by
163 extracting these pigments in a 90% alkaline acetone solution, measuring spectroscopic light
164 absorbance (Ultrospec 2100 Pro UV/Visible spectrophotometer, Amersham Biosciences, UK) at
165 665 and 430 nm wavelengths and converting these measurements into biomass using the
166 trichromatic equations by Wetzel and Likens (1979). Chl *a* content of periphyton was used as an
167 estimate of the total biomass of the community. The ratio of carotenoids and Chl *a* was
168 determined, as this may indicate heavy metal stress.

169 Protocols given by Pandey et al. (2014) was followed for estimating the intracellular
170 concentration of Cu, Zn and Pb. Periphytic algal samples were washed with 2mM EDTA
171 solution for 10 min and digested with concentrated HNO₃, H₂O₂ (80 %) and deionized water in
172 1:1:3 ratios on a hot plate at 90 °C (Bates et al., 1982). The residue was dissolved in 2% (v/v)
173 nitric acid and the final volume adjusted to 10 ml before measuring metal concentration with an
174 atomic absorption spectrophotometer (Perkin-Elmer, AAnalyst 800).

175 The number and volume of lipid bodies (LBs) were determined in 50 cells of a taxon as
176 per Pandey et al. (2015). Deformities (DFs) in diatom frustules were examined (in 500 frustules)
177 in fresh as well as through their permanent slide at 1000 x magnification as per Falasco et al.
178 (2009) and Pandey et al. (2014).

179 *Statistical analysis*

180 The Shannon index, species richness and Jaccard's index of periphytic diatom community was
181 estimated using "PAST" software (Natural History Museum, University of Oslo) (Hammer et al.,

2001). Data were statistically analyzed by one-way analysis of variance (ANOVA) followed by Tukey's HSD test for comparing various means.

3. Results

Physico-chemical characteristics of river water

Table S1 shows some important physicochemical characteristics of river water during three time periods. The current velocity showed tremendous fluctuations; it was maximum during the rainy season, but summer and winter season showed relatively lower current velocities. The rainy season not only had the highest flow rate, it also showed much greater fluctuations in current velocity. Insofar as the pH of the water is concerned, it was slightly above 7 in a majority of cases; no clear cut seasonal pattern was discernible. The conductivity was maximum during the summer season in comparison to the other two times of sampling. Nitrate-nitrogen occurred at higher concentration during summer in comparison to the other two seasons of sampling. Total phosphorus concentration also varied seasonally. It was the maximum during the rainy season followed in decreasing order by winter and summer. Dissolved silica content was around 10 mg l⁻¹ in most of the cases, but was maximum during the rainy season. The table makes it amply clear that low concentrations of heavy metals did not vary much in the river water.

Metal release rate from MDS and intracellular metal content

The rate of release of metal ions from MDS was studied in relation to time, and the data are presented in Figs. 2, S1 and S2. The rate of release of copper ions was a function of its concentration in MDS (Fig 2). Greater the concentration of metal ion inside MDS, greater was the rate of release of the test metal ion from MDS. Another important pattern has been the effect of time on the release of metal ions. The release of copper occurred very rapidly during the first

204 week and then it gradually declined with the passage of time and considerably lower rates were
205 seen in the third and the fourth week of sampling (Fig. 2). The initial curve was steeper at
206 the highest considered concentration of copper in MDS. Another important pattern evident in the
207 experiment has been marked influence of season on the rate of release of copper ions. It was the
208 maximum during the rainy season followed in decreasing order by summer and winter seasons.
209 The rate of release of zinc could be found in Figure S1. the general pattern of the rate of release
210 was similar to that of copper. It was maximum during rainy season and there was a clear-cut
211 effect of the time duration on the rate of release. The release rate of lead was low relative to the
212 release rates for copper and zinc (Fig. S2); however, the general pattern showed the effect of
213 concentration and time.

214 Intracellular accumulation of the test metals by the algal periphyton was also studied and
215 the data could be found in Figs. 1, S1 and S2. Intracellular concentration of copper increased
216 concomitantly with increase in concentration of copper in MDS (Fig. 1). It was maximum at the
217 highest tested concentration of copper and the lowest at the lowest tested concentration in MDS.
218 In almost all the cases, the intracellular level of copper attained maxima in the first week, and
219 thereafter it declined at all the tested concentrations of copper in MDS. Insofar as the effect of
220 season is concerned, maximum intracellular copper accumulation occurred in the summer season
221 and the minimum during the rainy and winter seasons. The intracellular accumulation of zinc
222 also followed a pattern (Fig. S1) which broadly matched with that obtained for copper. It
223 deserves mention that zinc was intracellularly accumulated much more in comparison to copper
224 and lead. There was clear-cut effect of metal concentration in MDS and time. The periphytic
225 community accumulated lower intracellular concentrations of lead than concentrations of copper

226 and zinc and this lower level of lead is consistent with the lower release rate of lead from the
227 MDS (compare Fig. S2 with Figs. 1 and S1).

228 *Periphytic community structure*

229 Periphytic growths could be seen on clay substrates placed in the river (Fig. 3).
230 Colonization of algae onto clay tiles started with the appearance of yellowish-cottony patches
231 intermixed with a few green patches which persisted for a time period of two weeks. The general
232 pattern of colonization remained similar in the control and metal-filled MDS. Periphytic biomass
233 on clay tiles at any point of time was a function of immigration, growth and loss due to
234 emigration or other factors. The periphytic colonization on the clay tiles was found quickest in
235 the summer season followed by winter and was the slowest during the rainy season. After two
236 weeks of colonization, green patches became more marked overshadowing the yellowish cottony
237 patches in the third week. In the fourth week all the substrates were covered with periphytic
238 growths of various colours ranging from yellow to green, blue green and brown.

239 After substrates were placed in the river, periphytic colonization was initiated by a
240 mixture of adnate diatoms and small coccoid green algae. Colonization in the control (without
241 metal) was visible to the naked eye after two days while MDS filled with metal solution,
242 depending on the concentration of metal, took a slightly longer time to attain the same stage. The
243 initiation of algal colonization was well marked by interspersed yellowish-green circular patches
244 of algae on the clay surface of MDS. Besides diatoms, filamentous green algae were also visible
245 with stalked and non-stalked diatoms colonizing MDS surface. Unicellular coccoidal green algae
246 were also seen in the periphytic algal community along with a few cyanobacterial cells.
247 Periphytic communities of summer, winter and rainy seasons were compared using Jaccard's
248 similarity coefficient, and cluster dendrograms were prepared (Fig. S3). The figure makes it clear

249 that the community of the rainy season was vastly different from that of the summer and winter
250 seasons. Summer and winter communities showed great resemblance with each other.

251 Fig. 4 depicts the cell density and total biovolume of the periphytic community during the
252 three seasons. Initially, during the first week of colonization, the periphytic community showed a
253 relatively smaller number of cells, which later on followed an increasing trend in the subsequent
254 weeks attaining the maximum value in the fourth week of the experiment. Accrual of algal
255 periphyton was the lowest during the rainy season and the maximum during the summer in the
256 case of the control substrates. The cell density was the highest in the case of the control
257 substrates but declined in the case of periphyton developing on the metal-filled MDS. Whereas
258 copper and zinc caused substantial toxic effects, lead was the least toxic. There was a distinct
259 concentration-dependent effect of the test metals on the cell density as also the total biovolume
260 of the periphytic community. Maximum inhibition of the cell density and total biovolume
261 occurred at the highest tested concentration of the metal, and the minimum at the lowest tested
262 concentration. It is interesting to note that the effect of metal concentration was particularly
263 marked in the first two week of colonization. However, it became less prominent during the
264 latter part of the experiment. In almost all the cases, especially from the third week onwards, the
265 cell number and biovolume of the periphyton of the control and various metal treatments do not
266 appear to be much different from each other. This was not the case with lead, as the communities
267 developing on lead diffusing substrates resembled with the control from the second week
268 onwards. Fig. 4 also reveals that the inhibitory effect of the test metals on the two parameters as
269 lesser during the rainy season in comparison to summer and winter seasons.

270 The periphytic assemblage developing on the control and metal-diffusing substrates
271 comprised individuals belonging to three major groups: Bacillariophyta (diatoms), Cyanophyta

272 (which are now preferred to be called as Cyanobacteria) and Chlorophyta (green algae) (Table
273 S2). Fig. 5 shows relative abundance of these groups in the assemblage after 7 and 28 days of
274 metal exposure, as also in the control assemblage for the same time period. The members of
275 Bacillariophyta dominated the periphytic assemblage in the control as well as various treatments.
276 Cyanobacteria became particularly abundant during the summer season. Green algae and
277 cyanobacteria became less abundant during the rainy season; on the other hand, diatoms were
278 most abundant (relative abundance > 80%) during this time period. Cyanobacteria responded in a
279 definite manner to metal enrichment. The relative abundance of cyanobacteria declined in metal-
280 filled MDS. The declination was concentration-dependent; it was greater at the highest tested
281 concentration of various test metals. It was particularly marked in the case of the one week old
282 community. Although the decline in cyanobacterial relative abundance by metal treatment was
283 not marked after 28 day exposure to the test metals, the relative contribution of cyanobacteria
284 was the greatest in the last week (28 days) of the experimental period in the case of the control
285 also. In a majority of cases, in the control and various treatments, the relative abundance of
286 diatoms slightly decreased during the later part of the experiment. During the initial stage of the
287 experiment (7 days), the relative proportion of green algae was slightly increased on MDS
288 releasing metal ions. However, it became almost similar to that of the control during the later
289 part of the experiment.

290 Species richness and Shannon diversity of the control and metal-exposed periphytic
291 community could be found in Fig. 6. The data have been presented for two time periods, i.e., 7th
292 and 28th day, for the three seasons because presenting all the data was considered cumbersome
293 and not necessary. Species richness was maximum in the case of the community growing on the
294 control substrates. The control substrates showed variations in species richness in the three

295 seasons. It was the minimum during the rainy season and the maximum in the summer. Species
296 richness of the community declined with increase in the level of metal enrichment in each of the
297 season. This particular effect was evident in the first week. In the case of the 28-day old
298 periphytic community developing on metal diffusing substrates, the effect of concentration was
299 not marked as the communities developing at various concentrations of a test metal and the
300 control had almost similar species richness. The declination of species richness was substantial in
301 the case of copper and zinc, but very mild in the case of lead exposed communities.

302 Species diversity also showed a pattern which showed decline, in comparison to the
303 control, dependent on the concentration of metal being released. Like species richness, Shannon
304 diversity also declined during the rainy season. The concentration-dependent decline in species
305 diversity was not marked in 28-day old periphytic community. Hence, the general pattern for
306 species diversity and species richness closely match each other.

307 Figs. S4 to S6 show relative abundance of common taxa in the 14-day old community
308 encountered on the control and various metal treatments during the three seasons. Fig. S4 shows
309 that the relative abundance of *Achnanthes exigua* was slightly enhanced by metal enrichment;
310 higher the concentration of metal, greater was the increase in relative proportion of this particular
311 species. The response was not metal specific; it was rather concentration dependent.
312 *Achnanthidium minutissimum* relative abundance increased following increase in metal
313 concentration. *Cymbella cymbriformis* showed mixed response. Its relative abundance initially
314 increased following low copper and zinc enrichment; however, decrease could be observed
315 subsequently. Lead could not elicit this kind of effect. The relative abundance of *Fragilaria*
316 *capucina* also increased with increase in the level of the test metals released by the substrates.
317 Lead was the least toxic to this particular diatom species.

318 *Gomphonema parvulum* was another diatom whose relative abundance increased
319 following metal treatment (except Pb exposure) (Fig. S5). In the case of copper and zinc
320 releasing substrates, the increase was evident only at the lowest tested concentration of these test
321 metals. Its relative abundance was very low during the rainy season and moreover metal
322 enrichment further reduced it. *Navicula recens* showed great sensitivity to the test metals. Its
323 relative contribution declined with increase in concentration of metal being released by the
324 substrates. *Nitzschia linearis* was another diatom that showed tolerance to the test metals as its
325 relative abundance increased with increase in the amount of metal ions released by MDS.
326 *Pinnularia conica* completely disappeared from the community during the rainy season, although
327 it was present at other times. This diatom also showed some tolerance to copper and zinc as
328 evident by enhancement of its relative abundance. However, lead did not exert significant on its
329 relative abundance.

330 *Ulnaria ulna*, another common large sized diatom species, showed a pattern (Fig. S6)
331 broadly resembling that of other metal sensitive species. Its relative abundance consistently and
332 gradually declined with increase in concentration of metal in the medium. *Chlorella vulgaris*
333 responded positively, although slightly to metal enrichment as was evident by increase in its
334 relative abundance especially during winter and summer months. The relative abundance of this
335 alga during the rainy season did not show any specific pattern; it remained almost similar but
336 considerably lower than at other time periods. The relative abundance of *Scenedesmus*
337 *quadricauda* fluctuated slightly, but it increased in several instances of metal treatment.
338 *Chroococcus limneticus* was slightly enhanced by low concentrations of metal ions; but higher
339 concentrations of the test metals were inhibitory. Its relative abundance was greatly decreased in
340 all the cases, including control, during the rainy season.

341 *Pigments and oil globules*

342 The amount of chlorophyll *a* and the ratio of carotenoids to chlorophyll *a* of the periphytic
343 community are presented in Table 1. The table makes it amply clear that metal stress (except Pb
344 exposure) reduced chlorophyll *a* content of the periphytic community. The reduction was strictly
345 concentration dependent. Zinc and Cu caused marked inhibition of chl *a* content of the
346 community; however, Pb did not affect chlorophyll content of the community. Insofar as the
347 ratio of carotenoids to chlorophyll *a* is concerned, both Cu and Zn brought about its enhancement
348 which was dependent on the concentration of metal being released by the substrate. The greater
349 the concentration of metal, the higher was the ratio of chlorophyll *a*: carotenoids. Lead did not
350 bring about appreciable change in the ratio of these two pigments.

351 In diatom species, oil globules (also referred to as lipid droplets) were observed (Fig. 7);
352 they were fewer in the case of the control. However, their number and size (% biovolume
353 relative to that of the entire cell) substantially increased under Cu stress (Fig. 7; Table S3). This
354 phenomenon was regularly observed in various diatom species under Cu and Zn stress during the
355 three seasons; however, a regular and consistent trend in relation to the concentration of metal in
356 MDS was observed only in eight of them, namely, *Achnanthes exigua*, *Navicula gregaria*,
357 *Navicula recens*, *Nitzschia amphibia*, *Nitzschia palea*, *Fragilaria capucina*, *Pinnularia conica*
358 and *Ulnaria ulna* (Table S3). In many of the species listed in the table, the number of oil
359 globules did not change considerably. However, tremendous fluctuations in biovolume of oil
360 globules occurred under metal stress. It is particularly interesting to note that the relative
361 contribution of oil globules to total cellular biovolume was as much as 70% in some cases of
362 metal treatment of *Achnanthes exigua*. At the other extreme is *Ulnaria ulna*; the control cells did
363 not show any oil globule, but metal treatment triggered the formation of small oil globules.

364 *Nitzschia linearis* also did not show any oil globule in the control; however, they did appear
365 under copper stress. Lead was not efficient in inducing the formation of oil globules.

366 *Deformities*

367 Deformities in diatoms frustules were examined in the control and metal treatments in the three
368 seasons (Table 2). In the three seasons, in comparison to the control, higher % deformed
369 frustules were examined under Cu, Zn and Pb treatments, which was also found to be
370 concentration dependent . However, under Cu and Zn treatments % deformities were more
371 prevalent than under Pb treatments.

372 **4. Discussion**

373 Prompted by earlier studies on nutrient diffusing substrates (Fairchild and Lowe, 1984; Pringle
374 and Bowers, 1984) and the preliminary work of Arnegard et al. (1998), a unique metal diffusing
375 porous clay substrate was developed which was cheap and ensured in situ study of the effect of
376 metal enrichment on periphyton (Pandey and Bergey, 2018). The substrate could very well
377 release metal ions throughout the period of the experiment; the rate of release was very high in
378 the beginning owing to large difference in concentration of metal inside MDS and water which
379 facilitated rapid diffusion of metal ions from the clay surface. But the diffusion of metal ions
380 from the porous clay surface considerably slowed down with the passage of time. This seems to
381 have primarily happened due to lowering of concentration of metal inside, although clogging of
382 some of the pores on the clay surface cannot be ruled out. A similar pattern of release of nutrient
383 ions has been earlier observed by researchers who deployed chemical diffusing substrates in
384 natural waters to study the effects of nitrogen and phosphorus enrichment on periphytic algal
385 communities (Scott et al., 2009). Of the three metals selected for the study, Cu and Zn could
386 easily diffuse out of the substrate, whereas Pb showed an extremely low rate of diffusion. This

387 variability may be related with greater effective radius of divalent ionic form of Pb (119 pm)
388 than that of Cu (73 pm) or Zn (74 pm). The rate of release of Cu, Zn and Pb ions from MDS
389 increased with their concentration in the MDS. The rate of release was the greatest during the
390 rainy season coinciding with the maximum current velocity during this period. Apparently, high
391 current velocity quickly swept away metal ions which were released by the substrate so as to
392 facilitate the release more of them from the surface during the rainy season.

393 Periphytic algae were able to colonize the substrate following a pattern which broadly
394 resembled with that obtained by other researchers (Steinman and McIntire, 1990; Pandey et al.,
395 2014). Metal diffusing substrates showed a similar pattern of colonization to control substrates;
396 however, the pace of colonization was slower on metal exposed substrates. The test metals
397 released by the substrates were taken up and intracellularly accumulated by periphytic
398 organisms. The rate of release of metal ions was maximum in the first week and therefore
399 greatest intracellular accumulation was registered during the first week. The release of metal ions
400 slowed down in subsequent weeks and this led to reduced transport of metal ions into the cell.
401 Concurrently, multiplication of periphytic cells also diluted the intracellular metal content with
402 the passage of time. The present observations showing intracellular accumulation of high
403 concentrations of the test metals by the periphytic community are in consonance with several
404 earlier researchers. For instance, Duong et al. (2008) and Morin et al. (2008a) noted
405 accumulation of high concentrations of cadmium by periphytic biofilm growing in metal
406 enriched environment. Similarly, Pandey et al. (2014) reported high intracellular accumulation of
407 heavy metals in the periphytic diatom community colonised on metal diffusing substrates
408 deployed in the river Ganges. Pandey and Bergey (2016) also found high build up of heavy

409 metals (Cu and Zn) in the periphytic diatom communities collected from metalliferous sites of
410 Rajasthan, India.

411 It has been pointed out by several researchers that metal content of biofilm can serve as a
412 reliable parameter for biomonitoring of metal concentration in water. (Newman et al., 1985;
413 Morin et al., 2008a). In most of the previous studies, metal content in water has been related with
414 total metal accumulated by periphytic biomass. This approach may not be very appealing
415 because in metal toxicity studies it is the intracellular metal which is responsible for toxic effects.
416 Behra et al. (2002) related metal content in water with intracellular metal content in periphytic
417 biomass. The present data are also based on intracellular metal content. In the present study, the
418 intracellular metal content, although increased initially in the first week, declined in the later part
419 of the experiment. This relates very well to concomitant decrease in the release of metal ion by
420 MDS. Arini et al. (2012a) similarly noted decline in metal content of metal-loaded periphytic
421 biofilm that were transferred from metal contaminated to the reference site. In a mesocosm study
422 also Morin et al. (2008b) could establish correlation between cadmium accumulation and
423 dissolved cadmium concentration.

424 The present study showed deleterious effects of the test metals on periphytic community.
425 The cell density and biovolume of the community followed a decreasing trend with increase in
426 concentration of metal being released by the clay substrates. Even chlorophyll *a* content per unit
427 area showed a metal concentration-dependent decreasing trend. The present observations are in
428 agreement with previous studies on metal toxicity to algal communities. For instance, Nirmala
429 Kumari et al. (1991) found algal abundance to be inversely related with concentration of metal
430 ions in a river system. Furthermore, Hill et al. (2000) noted decrease in concentration of
431 chlorophyll *a* following increase in metal level in a river impacted by mining operations. Zn and

432 Cd elicited more deleterious impact on chlorophyll a content of younger film than the older one
433 (Ivorra et al., 2000). Many other workers have also reported reduced periphytic biomass at
434 elevated concentrations of metals in stream mesocosms (Sigmon et al., 1977; Hedtke, 1984).
435 Conversely, de la Peña and Barreiro (2009) found pollutant load to be weakly related with
436 chlorophyll, and not at all related with the biomass. Metal pollution could not significantly alter
437 total standing crop of the phytoplankton community of a river (Montiero et al., 1995). Whitton
438 and Kelly (1995) and Stevenson and Pan (1999) considered biomass to be an undependable
439 criterion for the assessment of water quality. Notwithstanding various arguments made by
440 previous researchers, as mentioned above, the biomass parameters clearly reflected metal
441 concentration in the present study. However, some variation of results may be due to the
442 development of a periphyton community that is tolerant to the local metals concentration (i.e.,
443 due to a change in species composition).

444 In the case of the periphytic assemblage developing on the control substrates,
445 cyanobacteria were the most abundant during summer, which is characterized by high
446 temperature and light intensity. This observation agrees well with several previous reports of
447 high temperatures and optimal growth of cyanobacteria (Robarts and Zohary, 1987; Paerl, 1988).
448 In fact, cyanobacterial blooms have frequently been encountered in summer from many parts of
449 the world (Paerl, 1988). In the present study, the organisms belonging to cyanobacteria showed
450 greater sensitivity to the test metals compared to organisms belonging to other groups. On the
451 other hand, green algae slightly increased in relative abundance following metal enrichment. The
452 apparent increase in green algae may have been an artifact of using relative abundance because
453 of reductions in other groups and less impact on green algae (but possibly no actual increase).
454 Diatoms as a group showed intermediate response. Hence the present observations agree well

455 with Singh and Rai (1990) who noted the following order of Zn sensitivity to phytoplankton:
456 Cyanophyta > Chlorophyta > Bacillariophyta. Takamura et al. (1989) also observed greater
457 sensitivity of cyanobacteria to Cu, Cd and Zn, whereas green algae were relatively tolerant to the
458 test metals. Nirmala Kumari et al. (1991) in a study of phytoplankton of metal contaminated
459 river reported the following order of metal tolerance in algal groups: Chlorophyceae >
460 Bacillariophyceae > Cyanophyceae > Euglenophyceae. Whitton (1970) and Foster (1982) also
461 observed green algae, especially the members of Ulotrichales, remarkably tolerant to heavy
462 metals. On the contrary, Corcoll et al. (2012) found cyanobacteria to be tolerant to zinc in a
463 microcosm experiment, and Ivorra et al. (2000) and Kumar et al. (2012) also noted great
464 tolerance of cyanobacterial mats to metal pollution. Genter et al. (1987) noticed greater tolerance
465 to Zn in green algae than in diatoms and shifts in algal community from diatoms to filamentous
466 green algae and then to unicellular green algae with elevation of concentration of zinc.

467 Development of tolerant communities may also explain some of the differences in results
468 among studies. Major changes to community were reductions in species richness and diversity
469 (Shannon index). The effect showed concentration dependence. This obviously happened due to
470 disappearance or reduction in relative abundance of metal sensitive species from the community.
471 Niyogi et al. (2002) also reported a negative relationship between diatom diversity and stress
472 imposed by acid mine drainage. de la Peña and Barreiro (2009) studied the impact of abandoned
473 mine drainage on water quality and periphyton of nearby streams. Severely polluted sites had
474 diatom assemblages with lowered richness. Morin et al. (2008a, b) were able to relate Shannon
475 diversity index with the extent of pollution. Arini et al. (2012a, b) observed reduction of species
476 diversity of diatoms. On the contrary, Hirst et al. (2002) studied variations in metal concentration
477 and diatoms in 51 streams located in metal-mining areas of Wales and Cornwall, U.K. but could

478 not observe significant relationship between species diversity, species richness and evenness
479 with metal concentration in streams. Similarly, de la Peña and Barreiro (2009) observed higher
480 species diversity at moderately polluted site in comparison to the reference site. However, a
481 majority of earlier workers have reported decrease in number of species and reduced diversity of
482 metal exposed algal communities (Sharipova et al., 2007; Pandey et al., 2014, 2015; Pandey and
483 Bergey, 2016).

484 As already pointed out above, metal enrichment had tremendous impact on the
485 components of periphytic community; some species showed tolerance whereas others were
486 sensitive to the stress. *Achnantheidium minutissimum*, one of the abundant species in the
487 community, showed remarkable tolerance to the test metals. Its relative abundance increased
488 with increasing release of the test metals from the substrate. *A. minutissimum* is one of the few
489 species that colonizes a bare surface and is considered to be a good indicator of habitats
490 disturbed by physical extremes or by pollutants (Rott, 1991). A majority of earlier studies show
491 relative abundance of this species at elevated concentrations of metals in water (Cattaneo et al.,
492 2004; Luís et al., 2011; Arini et al., 2012b; Cantonati et al., 2014). Yoshiaki et al. (2004) showed
493 increased relative abundance of *A. minutissimum* with rising concentrations of copper, lead and
494 zinc, and concluded that it was tolerant to metals due perhaps to its smaller size. However, Luís
495 et al (2009) considered it to be neutrophilous and avoided high metal concentrations. On the
496 other hand, a few researchers believe it to be sensitive to heavy metals (Sabater, 2000; Blanck et
497 al., 2003). *A. minutissimum* can tolerate large variations in pH (Verb and Vis, 2000; Luis et al.,
498 2011) and grows well at high current velocities (Duncan and Blinn, 1989). Interestingly, Sgro et
499 al. (2007) regarded this diatom as an indicator of clean water. It has been pointed out that *A.*
500 *minutissimum* and other small diatoms, firmly attached to the substrate, are often entrapped in

501 organic matrix which may impede the passage of ions, including metal ions, into the frustule
502 thereby protecting them from metal toxicity (Burkholder et al., 1990). In the same context, it
503 needs to be mentioned that *A. minutissimum* is a species complex, which showed morphological
504 and ecological variations (Potapova and Hamilton, 2007), as a result of which differences in
505 metal tolerance might also occur. *Achnanthes exigua* was another diatom which was found
506 tolerant to the test metals. Ruggiu et al. (1998) studied diatom remains in cores taken out from
507 sediments of a subarctic Italian lake and found *Achnanthes* spp. to be tolerant to metal pollution.
508 Hill et al. (2000) also observed the dominance of *Achnanthes* at a river site most impacted by
509 mining operations. *Cymbella cymbriiformis* showed tolerance to low and medium tested
510 concentrations of the test metals. Roch et al. (1985) found it to be common in a lake impacted by
511 mining activities.

512 *Fragilaria capucina* was another diatom slightly tolerant to the test metals. Pandey et al.
513 (2015 and 2016) also reported dominance of *F. capucina* under Zn exposure (field and
514 laboratory conditions) but not under Cu stress. Conversely, Bere and Tundisi (2012) reported *F.*
515 *capucina* to be sensitive to metal pollutants in a laboratory mesocosm. *Gomphonema parvulum*
516 also showed tolerance to the test metals. The ability of *G. parvulum* to tolerate metal enrichment
517 agrees well with several previous researchers. In a laboratory grown phytoplankton community,
518 *Gomphonema parvulum* has been found to be zinc tolerant (Loez et al. 1995). Metal pollution
519 caused a shift in dominance of the phytoplankton community of a river in favour of *G. parvulum*
520 and some other taxa (Montiero et al., 1995). Duong et al. (2008) also found *Gomphonema*
521 *parvulum* to be cadmium tolerant. In fact, it was one of the dominant organisms in outflow from
522 a mining tailing dam (Sabater, 2000). On the contrary, Bere and Tundisi (2012) recently reported
523 it to be metal sensitive. *Pinnularia conica* also showed metal tolerance in the present study. Gold

524 et al. (2002) observed *Pinnularia* sp. at sites containing high concentrations of cadmium and
525 zinc; several other researchers have found *Pinnularia* spp. to be tolerant to heavy metals (Hirst et
526 al., 2002; Pandey et al., 2014; Pandey et al., 2016). Absence during the rainy season was another
527 noteworthy feature of *P. conica* observed in the present study. Since rainy season had high
528 current velocity, this particular species may have poor colonization success in high flow. *Ulnaria*
529 *ulna* showed marked sensitivity to the test metals. Another notable feature of this taxon observed
530 during the present study was its sensitivity to high current velocity; it failed to establish during
531 the rainy season.

532 Among green algae, *Chlorella vulgaris* and *Scenedesmus* sp. showed tolerance to high
533 concentrations of test metals. Metal tolerance of these two taxa has been reported earlier by
534 several researchers. For instance, in a laboratory grown phytoplankton community, Loez et al.
535 (1995) found *C. vulgaris* to be zinc tolerant. Stokes et al. (1973) also found *Chlorella* growing in
536 a lake contaminated by smelter wastes to be metal tolerant. In a study of phytoplankton of river
537 Moosi, Nirmala Kumari et al. (1991) also reported *C. vulgaris* as a metal tolerant species. The
538 other green alga, *Scenedesmus quadricauda* has also been found to be copper tolerant growing
539 well in Portugal reservoirs receiving copper-enriched mine effluent (Oliveira, 1985). Pandey et
540 al. (2015, 2016) also found *Scenedesmus* sp. from chronically polluted water to be metal tolerant.

541 Diatoms accumulate a lot of lipids in their cell, and a large proportion of cellular volume
542 may be occupied by them in the form of lipid droplets, also referred to as oil globules (Pandey et
543 al., 2017). Keeping this in mind, the lipid droplets were enumerated and their volume determined
544 in the present study. The test metals induced the formation of lipid droplets in diatom cells;
545 higher the concentration of metal greater was the number and volume of lipid droplets. The
546 present observations are in agreement with several other researchers who noted greater lipid

547 accumulation in diatoms and algae under metal (Pandey et al., 2015; Pandey and Bergey, 2016;
548 Gautam et al., 2017; Pandey et al., 2017) and diverse environmental stresses (Sharma et al.,
549 2012). The lipid droplets had the largest volume at the highest tested concentration of the test
550 metals thereby suggesting greater lipid accumulation by them. Since the change in number and
551 volume of lipid droplet showed distinct relationship with metal concentration, these two may
552 also be tested more vigorously for their application as a criterion in metal biomonitoring.

553 In the present study, morphological deformities in periphytic diatoms were more
554 prevalent under metal stress than the control, which was found to be in agreement with available
555 literature. For example, Pandey et al. (2014) reported significantly higher % deformed frustules
556 in the diatom community under Cu, Zn and Pb stress in the River Ganges and also found positive
557 and statistically significant relationship between % deformed frustules and different metals (Cu,
558 Zn and Pb) exposure. Cantonati et al. (2014) also reported deformities in *Achnanthydium*
559 *minutissimum* in the samples collected from different parts of Europe, Russia and America
560 impacted with heavy metals and considered deformities in this diatom species as a biomonitoring
561 tool for heavy metal contamination.

562 Published reports also showed that the different diatom metrics examined were act
563 together to counter metal stress in periphytic diatoms. For example, Pandey et al. (2016)
564 examined LBs and DFs together in diatom frustules (*Fragilaria capucina*, *Nitzschia linearis* and
565 *N. amphibia*) from metalliferous (Cu and Zn) sites of India. Gautam et al. (2017) reported LBs
566 and DFs together in the cells of *Gomphonema pseudoaugur* from metal (Pb and Se)
567 contaminated waterbodies of India. Under laboratory conditions, Pandey et al. (2015) reported
568 induction of DFs and LBs in the diatom cells (*Navicula gregaria*, *Nitzschia linearis* and *N.*
569 *amphibia*) under Cu and Zn stress.

570 This study also demonstrates the in situ effects of metal toxicity followed by recovery of
571 periphyton (using traditional algal parameters and non-taxonomical parameters) in the time
572 period of 28 days. Pandey and Bergey (2018) also examined more or less similar results of
573 periphyton recovery under similar sets of experiment conditions. Under stress, recovery response
574 of periphyton depends upon various factors such as, seasons, nutrients availability, production of
575 extra-polymeric substances (EPS) and emigration and immigration rates of algal cells (Stevenson
576 and Peterson, 1991; Pandey and Bergey, 2018). Short-life span, flexible life-history and good
577 dispersal abilities of periphytic algae also plays significant role in understanding recovery
578 response of periphyton (Steinman and McIntire, 1990). Thus, recovery response of periphyton
579 indicates not only monitoring of stable conditions but also for changing conditions, such as
580 monitoring recovery of aquatic ecosystems.

581 Finally, as shown by our study as well as those of other researches, we conclude that
582 changes in diatom assemblages, both taxonomic and morphological features, will be an excellent
583 and specific indicators of metal contamination. This study may also helpful in developing proper
584 restoration practices by using periphyton that helps in protecting the integrity of pristine fluvial
585 ecosystems.

586 **5. Conclusions**

587 The metal diffusing substrate designed in the present study is cheap, environment-
588 friendly and reliable, and could be used for in situ study of effects of metals and all other toxic
589 chemicals on periphytic community. However, the chemical, i.e., ion or molecule, whose effect
590 one intends to assess on periphyton should be small enough to diffuse out of the clay surface. In
591 the present study, the periphytic diatom community responded in a definite manner to metal
592 stress, which leads to taxonomical (cell density, biovolume, chl a, caro./chl a ratio, species

593 composition, and biodiversity indices), morphological (deformities) and physiological (lipid
594 bodies) alterations and metal diffusing substrates should be possible to employ it for
595 biomonitoring of waterbodies impacted by heavy metals.

596 **Acknowledgement**

597 LKP thanks the Head, Department of Botany, and the coordinator, Centre of Advanced Study in
598 Botany, Banaras Hindu University, for the necessary facilities. LKP thanks UGC and CSIR, New
599 Delhi for financial assistance. This work was also supported by the "UGC Start Up Grant (2019-
600 2020) (File No. 30-494/2019 (BSR))". We are grateful to Dr. J.C. Taylor (North-West
601 University, South Africa) for a generous gift of Pleurax.

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Table 1. Concentration of chlorophyll *a* and the ratio of carotenoids to chlorophyll *a* in periphytic community exposed to the test metals.

| Treatment | 7 days | | 28 days | |
|-----------------|--|---------------------|--|---------------------|
| | Chl <i>a</i> ($\mu\text{g cm}^{-2}$) | Carot./Chl <i>a</i> | Chl <i>a</i> ($\mu\text{g cm}^{-2}$) | Carot./Chl <i>a</i> |
| Control | 0.95±0.12 | 0.86±0.12 | 4.88±0.21 | 0.98±0.15 |
| Cu ^L | 0.85±0.21 | 1.00±0.14 | 4.81±0.22 | 0.99±0.24 |
| Cu ^M | 0.55±0.08 | 1.57±0.21 | 4.87±0.21 | 1.16±0.21 |
| Cu ^H | 0.15±0.02 | 1.86±0.25 | 4.85±0.25 | 1.21±0.22 |
| Zn ^L | 0.92±0.10 | 0.93±0.14 | 4.85±0.18 | 1.21±0.24 |
| Zn ^M | 0.67±0.08 | 1.02±0.08 | 4.84±0.15 | 1.11±0.25 |
| Zn ^H | 0.40±0.05 | 1.42±0.09 | 4.87±0.27 | 1.05±0.21 |
| Pb ^L | 0.95±0.13 | 0.93±0.11 | 4.85±0.16 | 1.00±0.18 |
| Pb ^M | 0.95±0.19 | 1.02±0.10 | 4.88±0.17 | 1.05±0.16 |
| Pb ^H | 0.95±0.20 | 0.95±0.13 | 4.90±0.16 | 1.04±0.14 |

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Table 2. Seasonal response pattern of attached algal community under in situ heavy metal enrichment expressed in terms of % deformity. Values given in the table represent the % deformities values on 7 and 28 days. Superscripts L, M and H denotes Low (1 g l^{-1}), Medium (2.5 g l^{-1}) and High (5 g l^{-1}) concentrations of Cu, Zn and Pb filled inside metal diffusing substrates. For more detail see figures 1,S1 and S2.

| Treatments | % Deformity | | |
|-----------------|-------------|-----------|-----------|
| | Winter | Summer | Rainy |
| Control | 0.02-0.07 | 0.02-0.09 | 0.05-0.11 |
| Cu ^L | 0.65-0.03 | 1.0-0.11 | 0.72-0.04 |

| | | | |
|-----------------|------------|-----------|-----------|
| Cu ^M | 1.68-0.21 | 2.1-.25 | 1.55-0.22 |
| Cu ^H | 2.45-0.32 | 3.0-0.22 | 2.72-0.15 |
| Zn ^L | 0.57-0.12 | 0.8-0.12 | 0.52-0.05 |
| Zn ^M | 1.83-0.30 | 2.0-0.14 | 1.92-0.22 |
| Zn ^H | 2.29-0.22 | 2.50-0.18 | 2.52-0.29 |
| Pb ^L | 0.18-0.04 | 0.21-0.05 | 0.15-0.03 |
| Pb ^M | 0.27-0.025 | 0.25-0.07 | 0.29-0.02 |
| Pb ^H | 0.42-0.05 | 0.35-0.09 | 0.49-0.07 |

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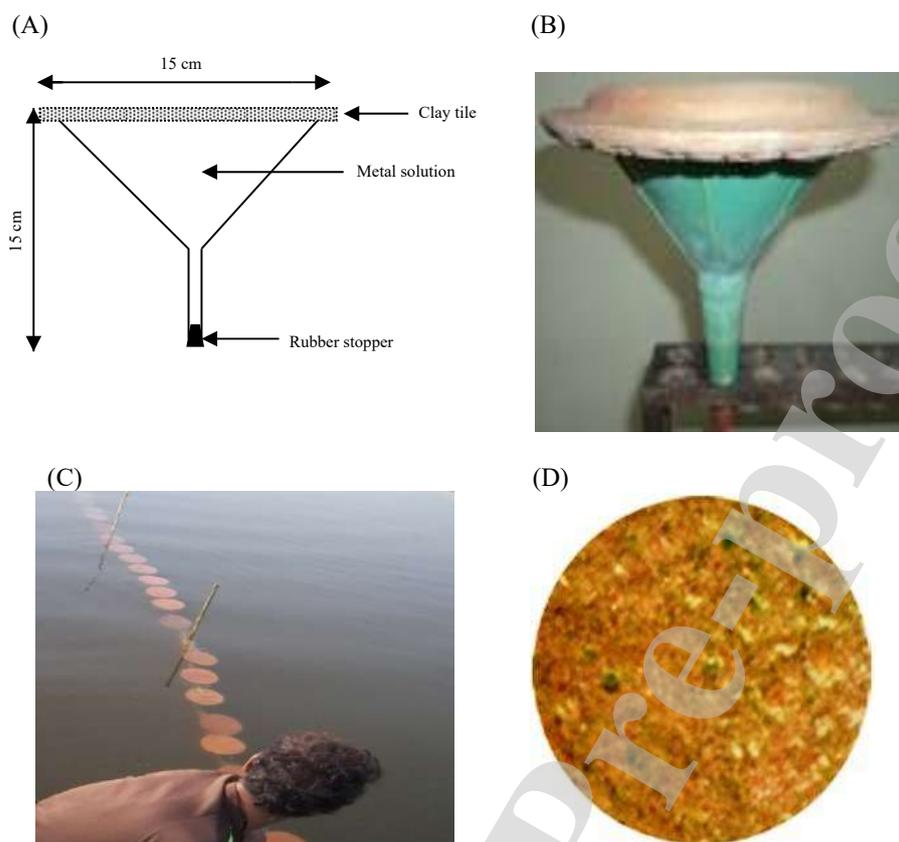


Fig. 1. (A) Diagrammatic representation of the design of metal diffusing substrate (MDS); (B) Photograph of an MDS; (C) Deployment of MDS in the river; (D) Periphytic colonization on the porous clay surface of MDS.

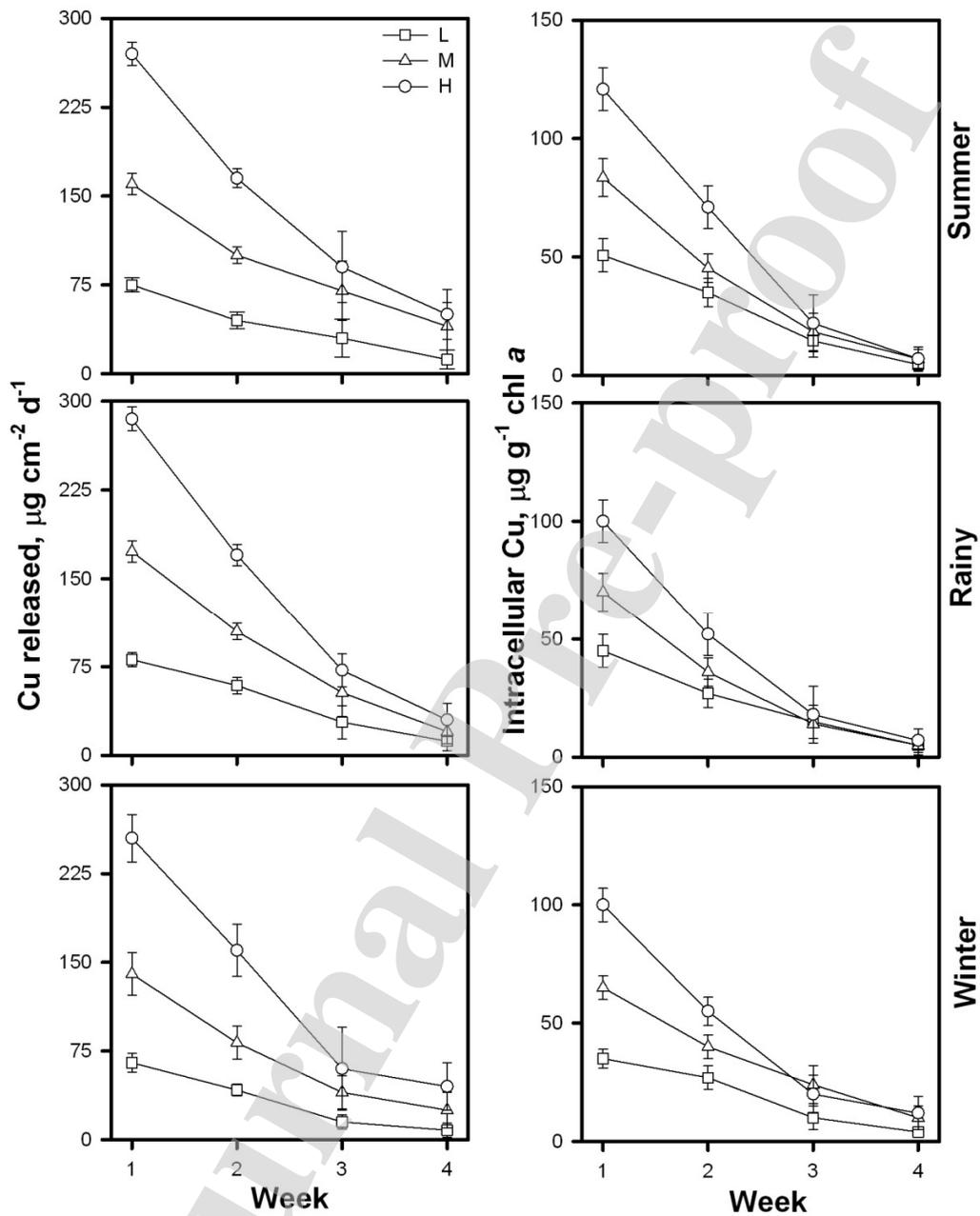


Fig. 2. Seasonal variations in the rate of release of Cu from MDS filled with low (L; 1 g l^{-1}), medium (M; 2.5 g l^{-1}) and high (5 g l^{-1}) concentrations of Cu, and intracellular Cu content in the periphyton. Vertical bars show standard error of means.

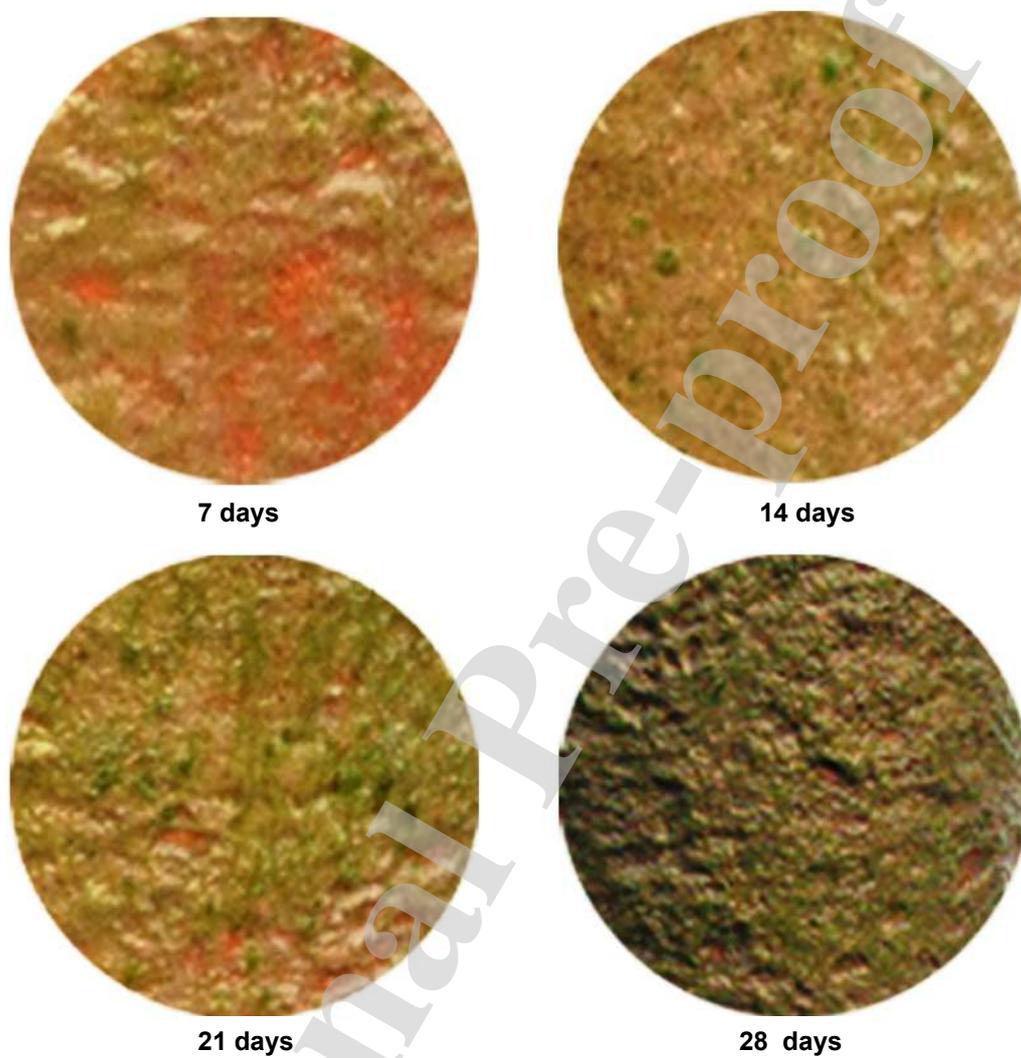


Fig. 3. The pattern of periphytic colonization onto clay surface of MDS. The period of the experiment was May to June, 2010.

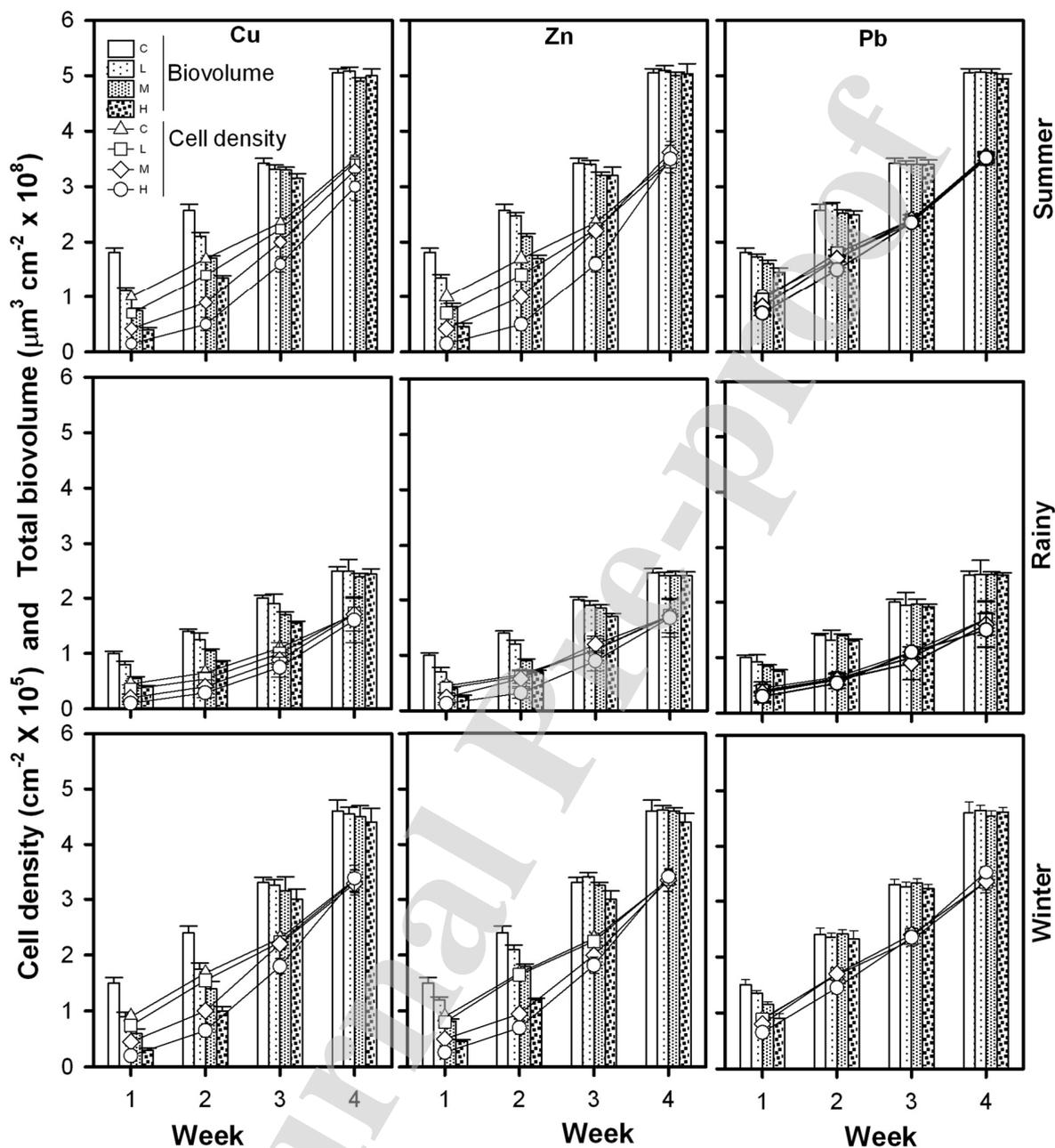


Fig. 4. Cell density and total biovolume of periphytic algal assemblages in the three seasons at low, medium and high concentrations (see Figs. 1, S1 and S2 for details) of the test metals. In the figure bars represent total biovolume while line plot represent cell density.

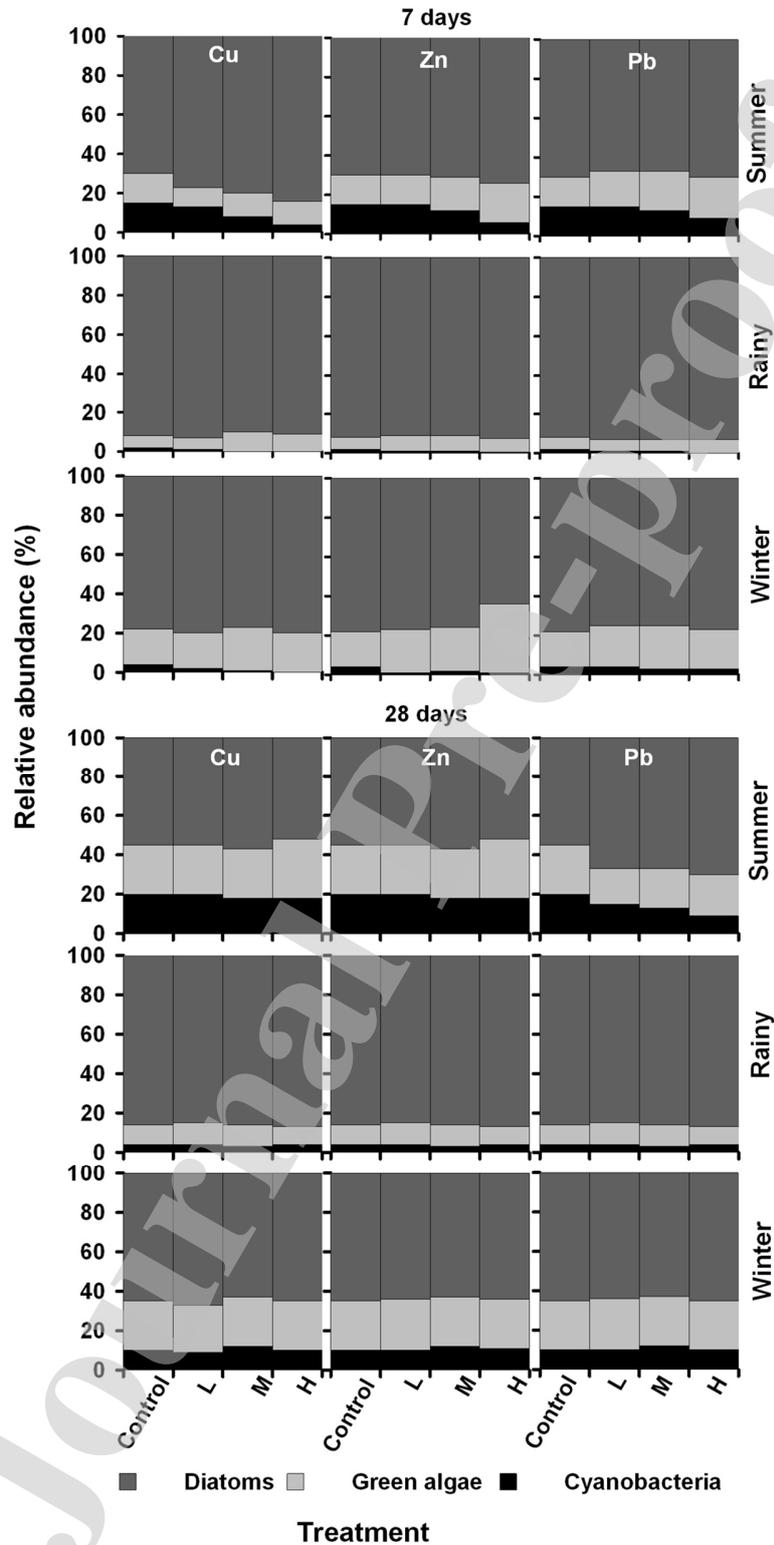


Fig. 5. Seasonal variations in the relative abundance of various groups in the periphyton at various concentrations of the test metals in MDS.

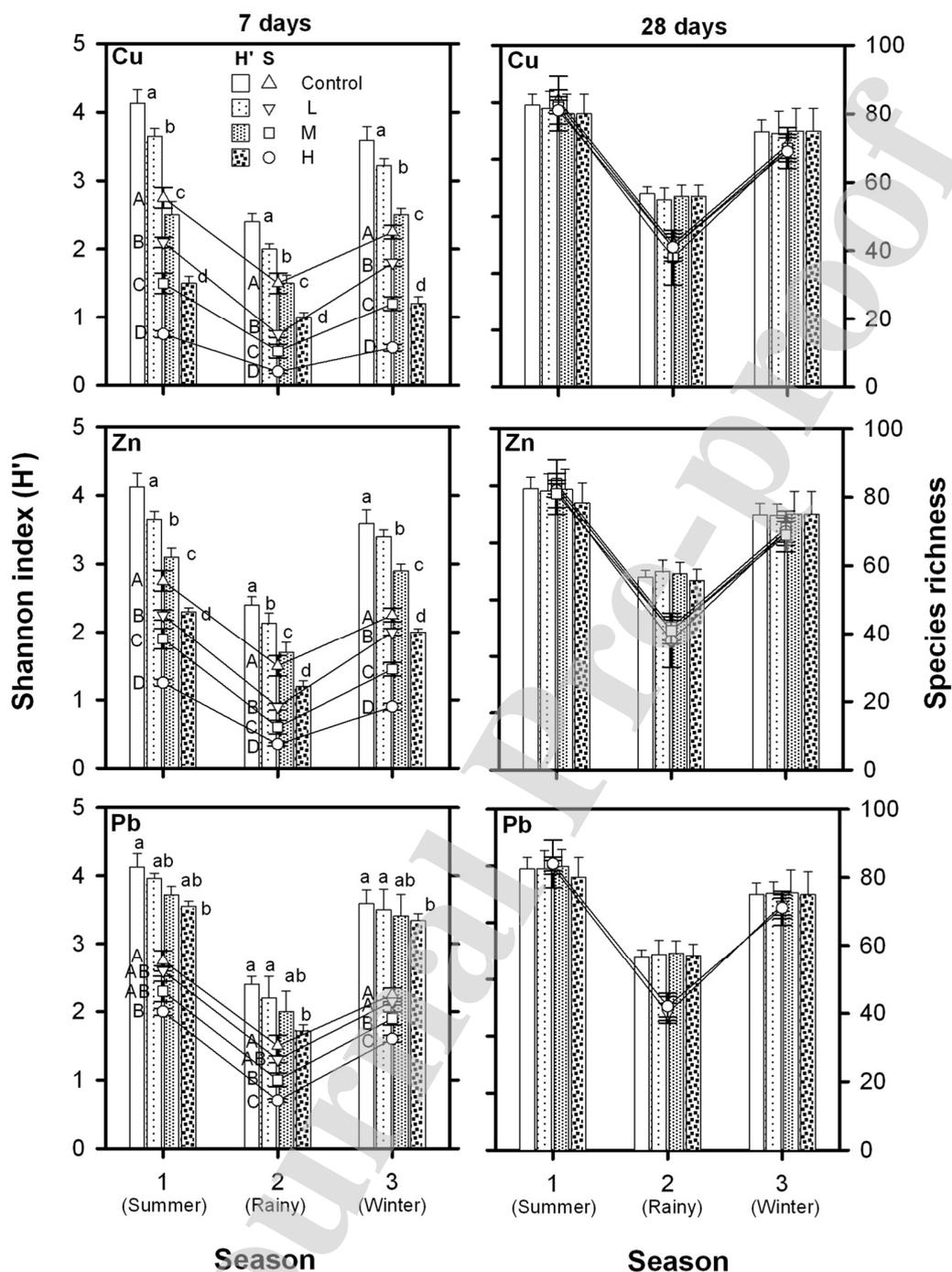


Fig. 6. Species richness and Shannon index (H') (species diversity) of the periphytic algal assemblage exposed to low, medium and high concentrations (see Figs. 1, S1 and S2 for details) of test metals for 7 and 28 days. Data bearing different letters for different seasons are significantly different ($p < 0.05$; Tukey's HSD test) from each other.

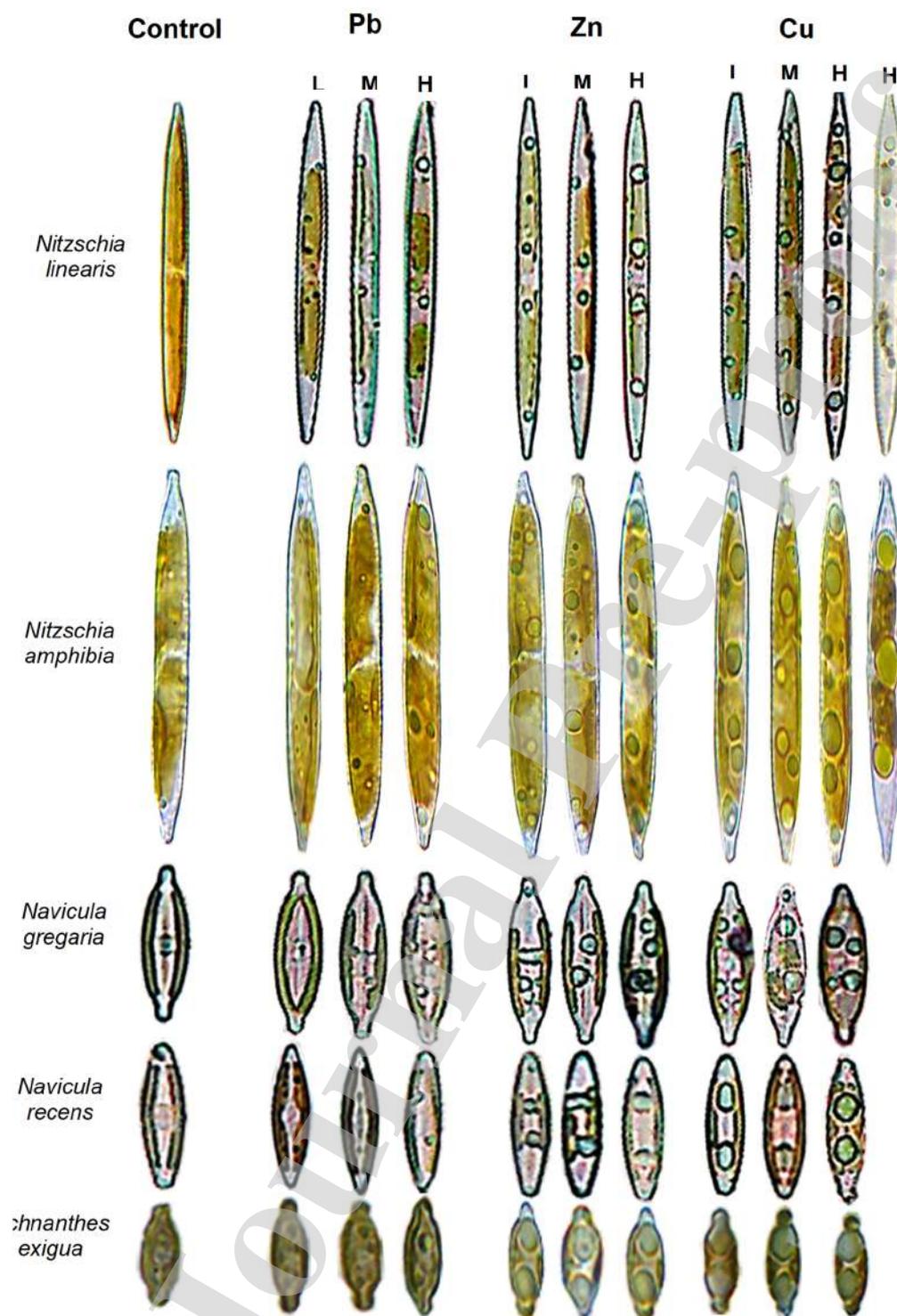
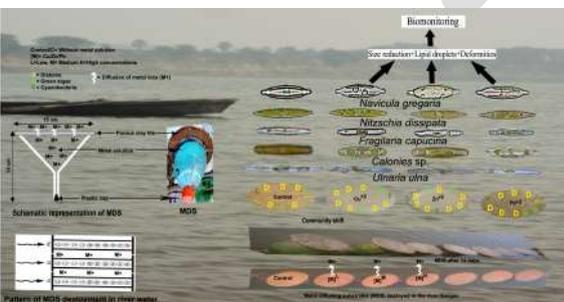


Fig. 7. Lipid droplets in live diatoms of the periphytic algal community in the control and metal treatments (L, M and H respectively represent low, medium and high concentrations of test metals; see Figs. 1, S1 and S2 for details).

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Highlights

- Metal diffusing substrates tested Cu, Zn & Pb toxicity to riverine periphytic algae.
- Taxonomic algal and diatom parameters were impacted by metal stress.
- Metals induce lipid bodies and morphological abnormalities in periphytic diatoms.

Author contributions

Lalit Kumar Pandey: Conceptualization, Investigation, Methodology, Formal analysis, Writing - original draft, Writing - review & editing and Funding acquisition.

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Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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